Cell Line Designation: COLO 205 cells

AddexBio Catalog No. C0009018

Cell Line Description:

Disease: Dukes' type D, colorectal adenocarcinoma

Origin: This line was isolated in 1975 by T.U. Semple, et al. from ascitic fluid of a 70-year-old Caucasian male with carcinoma of the colon. The patient had been treated with 5-fluorouracil for 4-6 weeks before removal of the fluid specimen.

Species: Homo sapiens

Tissue: colon; derived from metastatic site: ascites

Properties: Mixed, adherent and suspension

Morphology: Epithelial

Patient: Male, Caucasian, 70 years of age

Complete Medium: AddexBio-formulated RPMI-1640 (C0004-01) + 10% FBS

Subculture Procedure: Subculture before reaching confluence at a ratio of 1:6, and incubate at 5% CO2; 37°C incubator for 5-7 min. Resuspend in complete medium.

Medium Renewal: Two to three times weekly.

Freezing Medium: Complete culture medium supplemented 5% (v/v) DMSO. Store at liquid nitrogen vapor phase.

Additional Information: Additional product and technical information can be obtained from the catalog references and the Addexbio Technical Information site at www.addexbio.com, or by email at customersupport@addexbio.com.

Biosafety Level: 1

Appropriate safety procedures should always be used with this material. laboratory safety is discussed in the following publication: Biosafety in Microbiological and Biomedical Laboratories, 5th ed. HHS Publication No. (CDC) 93-8395. U.S. Department of Health and Human Services, Centers for Disease Control and Prevention. Washington DC: U.S. Government Printing Office; 2007. The entire text is also available online at www.cdc.gov/od/ohs/biosafty/bmbl4/bmbl4toc.htm

Use Restrictions:

These cells are distributed for research purposes only. Addexbio does not recommend third party distribution of this cell line, as this practice has resulted in the unintentional spreading of contaminated cell lines.

Handling Procedure for Frozen Cells:

To insure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If upon arrival, continued storage of the frozen culture is necessary, it should be stored in liquid nitrogen vapor phase and not at -70°C. Storage at -70°C will result in loss of viability.

Safety Precaution:

Addexbio highly recommends that protective gloves and clothing always be used and a full face mask always be worn when handling frozen vials. It is important to note that some vials leak when submersed in liquid nitrogen and will slowly fill with liquid nitrogen. Upon thawing, the conversion of the liquid nitrogen back to its gas phase may result in the vessel exploding or blowing off its cap with dangerous force creating flying debris.

1. Twenty four hours before thawing the cells, a culturing flask has to be pre-coated with S1 described under media and solutions. For growth area of 75 cm^2, add 5 mL of the S1 and rock gently to allow complete coverage of the flask surface.

2. Incubate the freshly coated vessel(s) in a 37°C incubator overnight (it is preferable to use tissue culture vessels with tightened, plug-seal caps to prevent evaporation during the coating process). [If extra flasks are coated, store coated flasks with S1 at room temperature, light protected, up to 1 month. Suction off solution before plating cells.]

3. Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the O-ring and cap out of water. Thawing should be rapid (approximately 2 minutes).

4. Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions.

5. Transfer the vial contents to the centrifuge tube containing 9.0 mL complete culture medium and spin at approximately 125xg for 5 to 7 minutes. (Optional if one wants to completely remove DMSO)

4. Resuspend cell pellet with the recommended complete medium and dispense into a new culture flask. It is important to avoid excessive alkalinity of the medium during recovery of the cells. It is suggested that, prior to the addition of the vial contents, the culture vessel containing the complete growth medium be placed into the incubator for at least 15 minutes to allow the medium to reach its normal pH (7.0-7.6).

5. One may also transfer the vial contents into a new culture flask if removal of DMSO is not important. It is suggested that, prior to the addition of the vial contents, the culture vessel containing the complete growth medium be placed into the incubator for at least 15 minutes to allow the medium to reach its normal pH (7.0-7.6).

6. Incubate the culture at 37°C in a suitable incubator for 24-48 hours for cell attachment. A 5% CO_2 in air atmosphere is recommended.

References for COLO 205 cells:

1. Semple TU, Quinn LA, Woods LK, Moore GE. Tumor and lymphoid cell lines from a patient with carcinoma of the colon for a cytotoxicity model. Cancer Res. 1978 May;38(5):1345-1355.

Lot Specific Information Sheet for AddexBio Cat #: C0009018

Lot Number: 1495638

Designation: COLO 205 CELLS

Total Cells/mL: >1.1 x10^6

Expected Viability: 70.0-75.1%

Ampule Passage #: 10

Dilute Ampule Content: 1:10 (T-25) or 1:15 (T-75)

Volume/Ampule: 1 mL

A T-75 set up at a dilution of 1:15 reaches approximately 50% to 60% confluence in 3 days.

Lot Specific Information Sheet for AddexBio Cat #: C0009018

Lot Number: 1529353

Designation: COLO 205 CELLS

Total Cells/mL: >1.3 x10^6

Expected Viability: 77.2-81.5%

Ampule Passage #: 12

Dilute Ampule Content: 1:10 (T-25) or 1:15 (T-75)

Volume/Ampule: 1 mL

A T-75 set up at a dilution of 1:15 reaches approximately 50% to 60% confluence in 3 days.